



Priscila Gonçalves Calmeirão

[Licenciada em Engenharia Biomédica]

Energy production on a Microbial Fuel Cell with *Serratia marcescens*, using conductive fabric as electrodes

Dissertação para obtenção do Grau de Mestre em
[Engenharia Micro e Nanotecnologias]

Orientador: Isabel Ferreira, Professora Ph.D., UNL-FCT

Co-orientador: Carla Carneiro, Professora Ph.D., IPS-ESTS/UNL-FCT

Júri:

Presidente: Prof. Hugo Miguel Águas

Arguentes: Prof. Rui Amaral

Prof. Carla Carneiro



FACULDADE DE
CIÊNCIAS E TECNOLOGIA
UNIVERSIDADE NOVA DE LISBOA

Dezembro, 2018

“Energy production on a Microbial Fuel Cell with *Serratia marcescens*, using conductive fabric as electrodes”

Copyright © Priscila Gonçalves Calmeirão, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa.

A Faculdade de Ciências e Tecnologia e a Universidade Nova de Lisboa têm o direito, perpétuo e sem limites geográficos, de arquivar e publicar esta dissertação através de exemplares impressos reproduzidos em papel ou de forma digital, ou por qualquer outro meio conhecido ou que venha a ser inventado, e de a divulgar através de repositórios científicos e de admitir a sua cópia e distribuição com objectivos educacionais ou de investigação, não comerciais, desde que seja dado crédito ao autor e editor.

À minha família e amigos pela perseverança de não me deixar desistir.

Agradecimentos

Estes últimos anos foram uma jornada de enorme aprendizagem que estarei sempre grata pelo conhecimento que ganhei e pelas pessoas que conheci.

Apesar de ter sido um percurso complicado não posso deixar de agradecer aos meus orientadores Professora Doutora Carla Carneiro e Professora Doutora Isabel Ferreira pelo apoio mesmo perante alguma distância e falta de tempo da minha parte foram sempre prontos para resolver os problemas que apareciam.

Agradeço ao Professor Doutor José Moura pela oportunidade que me deu de pertencer ao seu grupo de investigação durante dois longos anos.

Pelo trabalho suportado pela Faculdade de Ciências da Universidade Nova de Lisboa (FCT-UNL) e pelo REQUIMTE/Centro de Química Fina e Biotecnologia da Universidade Nova de Lisboa.

Ao laboratório de Bioquímica e Microbiologia da Escola de Tecnologia de Setúbal - Instituto Politécnico de Setúbal (IPS-ESTS) por me ter dado a minha primeira base de formação nesta área e por me receber sempre de braços abertos.

Por todo o suporte e incentivo a minha família, que teve um papel muito importante na conclusão desta tese e que sempre me apoiou e acreditou em mim. Para a minha mãe e ao meu irmão o meu eterno agradecimento, por me fazerem a pessoa que eu sou hoje, muito obrigado aos dois.

Resumo

A produção de energia é um tema de tendência atualmente, com o excesso de combustíveis fóssil e a disseminação de energias renováveis. A busca por novas formas de produzir energia é ampla. Neste caso, uma bactéria gram-negativa *Serratia marcescens* é usada com uma célula de combustível microbiana (MFC). Este dispositivo irá converter a energia química proveniente da atividade catalítica da bactéria em eletricidade. Para otimizar o sistema de referência, a fim de aumentar a sua eficiência elétrica, é utilizada uma deposição de filme de grafeno sobre uma camada de contato condutiva, materiais condutores recentes feitos de tecido são alternativa ao habitual filme comum de crômio-ouro, para manter a neutralidade e evitar a difusão de oxigênio utiliza-se uma Membrana de Troca de Protões (PEM) ou uma membrana diferente de acetato de celulose. Essa estrutura permite uma captura mais profunda dos eletrodos livres da câmara anódica.

Palavras-chave: MFC; Grafeno; Tecido Condutor; PLA; *Serratia marcescens*.

Abstract

The production of energy is a trend subject nowadays, with overdo of fossil fuels and the spread of renewable energies. The search for new ways to produce energy is wide; in this case, a gram-negative bacteria *Serratia marcescens* is used with a Microbial Fuel Cell (MFC). This device will convert the chemical energy coming from the catalytic activity of the bacteria into electricity. To optimize the reference system, in order to increase the electrical efficiency, a graphene film deposition is used over a conductive contact layer, recent conductive materials made of fabric are an alternative to the common chromium-gold film, to maintain the neutrality and avoid the oxygen diffusion it is used a Proton Exchange Membrane (PEM) and a different membrane made of cellulose acetate. This structure allows a more in-depth capture of the free electrodes from the anode chamber.

Keywords: MFC; Graphene; Conductive Fabric; PLA; *Serratia marcescens*.

List of Tables

| | |
|---|----|
| TABLE 1 - LIST OF THE MFC BASELINE TESTS AND INTRODUCTION OF <i>S. MARCESCENS</i> | 10 |
| TABLE 2 – STANDARD STATISTICS FOR THE 4 POPULATIONS IN STUDY. | 20 |
| TABLE 3 - WELCH ANOVA RESULTS. | 21 |
| TABLE 4 - KRUSKAL-WALLIS RESULT..... | 22 |
| TABLE 5 - PROPOSED Nº1 CONCENTRATIONS TO REACH OPTIMAL CONDITIONS. | 24 |
| TABLE 6 - PROPOSED Nº2 CONCENTRATIONS TO REACH OPTIMAL CONDITIONS. | 25 |
| TABLE 7 - PROPOSED Nº3 CONCENTRATIONS TO REACH OPTIMAL CONDITIONS. | 25 |
| TABLE 8 - REFERENCE CHARACTERISTICS USED FOR THE Cr/Au FILM DEPOSITION IN THIS STUDY. (FERNANDES ET AL., 2014) | 26 |

List of Figures

| | |
|--|----|
| FIGURE 1 - SCHEMATIC DIAGRAM OF A TYPICAL TWO-CHAMBER MICROBIAL FUEL CELL. A- RECTANGULAR SHAPE; B- SQUARE MINIMIZE SHAPE. (DU, LI, & GU, 2007) | 4 |
| FIGURE 2 – SCHEMATIC OF THE FUNCTIONING REACTIONS OCCURRING ON A MICROBIAL FUEL CELL. (LOVLEY, 2006) | 5 |
| FIGURE 3 - REFERENCE MICROBIAL FUEL CELL STRUCTURE ADAPTED FROM (FERNANDES, ROCHA, & CARNEIRO, 2014). | 10 |
| FIGURE 4 - GROWTH CURVE FOR <i>SERRATIA MARCESCENS</i> . B) PRESENCE OF PLA TRIAL 1; C) PRESENCE OF PLA TRIAL 2 | 13 |
| FIGURE 5 - A) PHOSPHATE BUFFER TRIAL IN TWO SUB-CELL COMPARTMENTS; B) GLUCOSE TRIAL IN TWO SUB-CELL COMPARTMENTS | 14 |
| FIGURE 6 – RESULTS FOR GLUCOSE BASELINE (1)..... | 15 |
| FIGURE 7 – RESULTS FOR METHYLENE BLUE BASELINE (2)..... | 15 |
| FIGURE 8 – RESULTS FOR THE COMBO GLUCOSE + METHYLENE BLUE BASELINE (3); A – DRY-UP CONDUCTIVE FABRIC; B – CONDUCTIVE FABRIC PREVIOUSLY BATHED IN PHOSPHATE BUFFER SOLUTION. | 16 |
| FIGURE 9 – ALL BASELINES, THIS GRAPHIC DISPLAYS ONLY THE TWO BEST RESULTS FROM ALL THE TRIALS PERFORMED..... | 17 |
| FIGURE 10 - <i>SERRATIA MARCESCENS</i> GROWTH CURVE WITH TEST CONDITIONS THROUGH 30 HOURS. | 18 |
| FIGURE 11 – INTRODUCTION OF <i>SERRATIA MARCESCENS</i> (4) IN THE MFC TRIALS..... | 19 |
| FIGURE 12 - MFC TEST IN SERIES WITH ALL THREE COMPARTMENTS CONNECTED TO THE CIRCUIT..... | 23 |
| FIGURE 14 - MFC STRUCTURE WITH GRAPHENE AS AN ELECTRODE. ADAPTED (FERNANDES, ROCHA, & CARNEIRO, 2014). | 26 |
| FIGURE 15 - SKETCH OF THE PLA/S.M. BIOCOMPATIBILITY TEST (ERLEN MAYER WITH <i>S. MARCESCENS</i>)..... | 35 |
| FIGURE 16 - STANDARD SETUP USED FOR ALL THE MFC TESTS USING A RESISTOR OF 1 MΩ..... | 36 |

List of Acronyms

3D – Three Dimensional

Au – Chemical element for Gold

BOD - Biochemical Oxygen Demand

Cu – Chemical element for Copper

CNT – Carbon Nanotubes

MFC – Microbial Fuel Cell

Ni – Chemical element for Niquel

PC - Polycarbonate

PDMS - Polydimethylciloxane

PEM – Proton Exchange Membrane

PLA – Polylactic Acid

S.m – *Serratia marcescens* also abbreviated as *S. marcescens*

Content

| | |
|-------------------------------|-------------|
| AGRADECIMENTOS | V |
| RESUMO | VI |
| ABSTRACT | VII |
| LIST OF TABLES | VIII |
| LIST OF FIGURES | IX |
| LIST OF ACRONYMS | X |
| INTRODUCTION | 3 |
| MATERIALS AND METHODS | 9 |
| RESULTS AND DISCUSSION | 11 |
| CONCLUSION | 29 |
| FUTURE PERSPECTIVES | 31 |
| REFERENCES | 33 |
| APPENDIX 1 | 35 |

Introduction

1.1. Microbial Fuel Cell

A Microbial Fuel Cell (MFC) is a device that converts chemical energy, contained in the chemical bonds of organic compounds, to electrical energy through catalytic reactions of microorganisms under anaerobic conditions. The electrons produced by the bacteria from these substrates are transferred to the anode (negative terminal) and flow to the cathode (positive terminal) and their linked by a conductive material containing a resistor or operated under a load.

It has been known for many years that it is possible to generate electricity directly by using bacteria to break down organic substrates. Bacteria can be used in Microbial Fuel Cells to generate electricity while accomplishing the biodegradation of organic matters or wastes.

Figure 1 shows a schematic diagram of a typical MFC for producing electricity. The anodic and cathodic chambers are separated by a proton exchange membrane (PEM). Keeping the microorganisms separated from oxygen or any other end terminal acceptor other than the anode makes the generation of electric current possible, requiring only an anaerobic anodic chamber. A MFC mimics the way nature uses to recycle the energy from renewable biomass, it does not require fossil fuels, which makes it a clean energy. (Du, Li, & Gu, 2007)

The process is simple, microbes are injected into the anode of the MFC that oxidize when added a substrate and generate electrons and protons in the process, the electrons in the anode are transferred to the cathode through an external circuit, the protons cross over a PEM to move into the cathode, as in figure 2. The performance of an MFC is related to the electrochemical reactions that occur between the organic substrate, at a low potential, such as glucose and the final electron acceptor with a high potential, such as oxygen. Microbes in the anode will extract electrons and protons. In the dissimilative process of oxidizing organic substrates, however, it's not possible to

know the ideal cell voltage because the electrons are transferred to the anode from the organic substrate through a complex respiratory chain that varies from microbe to microbe and even for the same microbe when growth conditions differ. (Du, Li, & Gu, 2007) (Pant, Van Bogaert, Diels, & Vanbroekhoven, 2010)

Microbial cells not only can produce energy (Eq.3) but also have as products water (Eq.2), heat and, depending on the fuel source, little amount of gases release such as carbon dioxide (Eq.1), nitro dioxide and other. (Fernandes, Rocha, & Carneiro, 2014)(Kalathil, Patil, & Pant, 2017)

Anodic reaction: $\text{CH}_3\text{COO}^- + 2\text{H}_2\text{O} \rightarrow (\text{microbes}) 2\text{CO}_2 + 7\text{H}^+ + 8\text{e}^-$ Eq. (1)

Cathode reaction: $\text{O}_2 + 4\text{e}^- + 4\text{H}^+ \rightarrow 2\text{H}_2\text{O}$ Eq. (2)

Overall: $\text{CH}_3\text{COOH} + 2\text{O}_2 \rightarrow 2\text{CO}_2 + 2\text{H}_2\text{O} + \text{Biomass} + \text{Electricity}$ Eq. (3)

There are a few types of MFC system designs considering the disposition of the anode and cathode chambers. According to (Du, Li, & Gu, 2007) we can find a single-compartment MFC, a two-compartment MFC, a Up-flow mode MFC and a Stacked MFC.

The system used in this study is a two-compartment MFC (figure 1). The anode and the cathode chambers are separated/connected by a Proton Exchange Membrane, this system allows the protons to move across to the cathode chamber while blocking oxygen diffusion into the anode chamber. The shapes of each chamber can be whatever the user design it to be, the most simple and also the one of choice for this study was a square chamber, as shown in figure 1B.

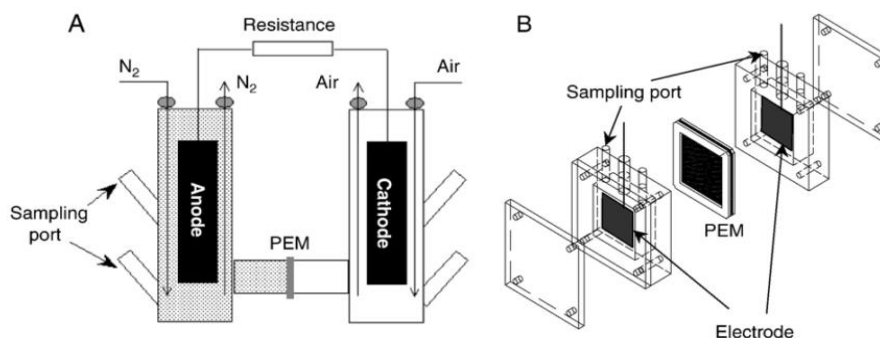


Figure 1 - Schematic diagram of a typical two-chamber microbial fuel cell. A- Rectangular shape; B- Square minimize shape. (Du, Li, & Gu, 2007)

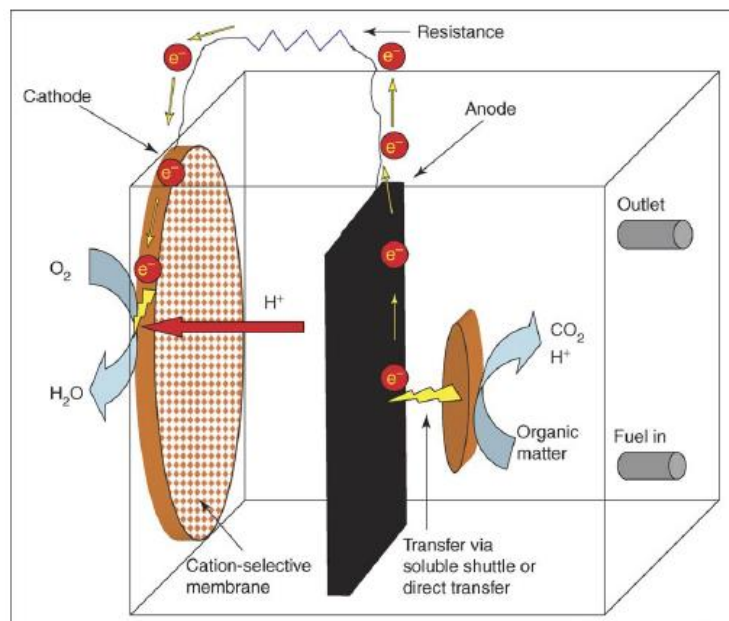


Figure 2 – Schematic of the functioning reactions occurring on a microbial fuel cell. (Lovley, 2006)

1.2. Materials for Microbial Fuel Cell Construction

A wide range of materials have been tested to improve the performance of MFCs. The performance of MFC directly depends on the kinetics of the electrode reactions within the cell, and its performance is very influenced by the materials they are made from. A superior biocompatible material will increase the bacterial adhesion and hence the life of the MFC. (Mustakeem, 2015)

Anodic materials must be conductive, biocompatible, and chemically stable in the reactor solution. Metal anodes must be noncorrosive stainless-steel mesh, but copper is not useful due to the toxicity to the bacteria. The most versatile electrode material is carbon, available as compact graphite plates, rods, or granules, as fibrous material (felt, cloth, paper, fibers, foam), and as glassy carbon. (Logan et al., 2006)

Cathode materials due to its good performance, ferricyanide ($K_3[Fe(CN)_6]$) is very popular as an experimental electron acceptor in microbial fuel cells. The greatest advantage of ferricyanide is the lower overpotential when using a plain carbon cathode, resulting in a cathode working potential close to its open circuit potential. The greatest disadvantage, however, is the insufficient re-oxidation by oxygen, which requires the catholyte to be regularly replaced or used in tests that required a shorter period of time.

The output power of MFCs is greatly constrained by the surface area of electrodes. The ohmic losses are directly proportional to the resistance of the electrode. A wider and effective surface area is considerably important to improve the efficiency of the cell. At

the same time, the interfacial impedance of the electrode material should be low to facilitate the electron transfer. At the cathode, the ionic conductivity is also required to facilitate triple phase boundary reaction. (Mustakeem, 2015)

1.3. Microbial culture: *Serratia marcescens*

The anodic electron transfer mechanism in MFC is a key issue in understanding the theory of how MFCs work. As mentioned above, microbes transfer electrons to the electrode through an electron transport system that either consists of a series of components in the bacterial extracellular matrix or together with electron shuttles dissolved in the bulk solution. (Du, Li, & Gu, 2007) Substrate is important for any biological process as it serves as carbon (nutrient) and energy source. The efficiency and economic viability of converting organic wastes to bioenergy depend on the characteristics and components of the waste material. (Ghanapriya, Rana, & Kalaichelvan, 2010) Theoretically, most microbes can potentially be used as a biocatalyst in MFC. In this study it was used *Serratia marcescens*, a gram-negative bacillus classified as a member of the *Enterobacteriaceae*, that has been recognized as a cause of hospital-acquired infection for the last two decades. *S. marcescens* can use a wide range of nutrients and has the ability to survive and grow under extreme conditions, including in disinfectant, antiseptics and double distilled water and it's easily recognized by its red colonies. (Hejazi & Falkiner, 1997)

1.4. Applications

The microfluidic MFC device developed could serve as a versatile platform for studying the interactive effects of cell growth, electrode reactivity, and ion transport at the micro-/Nano-scale. (Qian, Baum, Gu, & Morse, 2009)

Presently, most of the world's energy is derived from fossil fuels, but their efficiency, security and environmental effects hold it back from being a preferred energy resource. To meet future energy demands, there is a need of reliable, sustainable and clean energy source. While nuclear power may be an alternative mean of energy, a safer and reliable implementation is always a beneficial choice. Currently, real-world applications of MFCs are limited because of their low power density. (Pant. 2010)

There are a few applications being studied nowadays with Microbial Fuel Cells, the potential of producing energy out of microorganism opens a new perspective for the creation of Bioenergy, new alternative Bio-sensors and for the residual water's treatment. And the reason for that is that MFCs are theoretically energy profitable, based on their low energy consumption and direct electricity generation, they have a low adverse impact on the environment and, theoretically, have a good operational stability and low operational cost. (Liu, Cheng, 2014)

Wastewaters Treatment

A lot of energy is stored in wastewaters, capturing this energy is of great significance for meeting the world's energy needs, reducing wastewater handling costs and

increasing the sustainability of wastewater treatment. The microbial fuel cell can be used for electrical energy recovery from the organic pollutants in wastewaters. MFCs hold great promise for sustainable wastewater treatment. More importantly, MFCs are capable of directly producing electricity from the organic matter in wastewater with a high energy conversion rate, whereas the conversion of biogas (e.g., CH₄ or H₂) into electricity causes a significant energy loss of more than 60% (Rittmann, 2008).

Biosensors

According to previous studies (Kim, Chang, Gil, Park & Kim 2003) (Chang, Jang, Gil, Kim 2004) MFC could be used to continuously measure the biochemical oxygen demand (BOD) of wastewater for real-time monitoring. The directly proportional relation between the coulomb that is being produced from the microbial fuel cell and the strength of the wastewater make possible to use this technology as a BOD sensor.

Bio- Hydrogen

Hydrogen can be produced from certain forms of biomass by biological fermentation though yields are low. However by electrochemically expanding the cathode potential in a MFC circuit it is possible to directly produce hydrogen from protons and electrons produced by the bacteria. This technique will reduce the energy needed to make hydrogen directly from organic matter compared to that required for hydrogen production from water via electrolysis. Deriving the protons and electrons from organic matter instead of water, we can directly generate hydrogen at a low voltage using a type of MFC device that does not require oxygen. The use of a completely anaerobic process should increase the Coulombic efficiency of the MFC process using mixed cultures as oxygen diffusion through the cathode is avoided (Liu, Grot, Logan 2005).

Materials and Methods

This research is based on a previous work (Fernandes, Rocha, & Carneiro, 2014), consists on building a MFC with alternative materials such as polymeric structure and conductive fabric for electrodes.

First phase: redo a complete new MFC using the same design structure but replacing acrylic and glass parts with a PLA 3D printed piece, silicone was replaced with PDMS and Nafion PEM with cellulose acetate.

A compatibility test had to be done using PLA and the bacterial environment in use, *Serratia marcescens*, in order to make sure that the change of material wouldn't affect the bacteria growth. The process is displayed in figure 14 in Appendix 1.

Second phase: Replace from the original cell only the glass material with PC parts with five times the thickness of the glass, to make a less fragile and a more tightened cell. The electrodes were also replaces for [Cu+Ni+Au] conductive fabric from METALINE, instead of the Cr/Au + Carbon Nanotubes and Nafion PEM (Nafion N-117 membrane, 0.180mm thick, => 0.90 meq-7g exchange capacity)

Third phase: Introduction of graphene, in this phase two experiments were to be done (1) switch the carbon nanotubes for graphene using the same reference method; (2) combining graphene with the conductive fabric using the method performed in phase two of this research.

Since the experiment uses liquid solutions, it was mandatory that this new structure would avoid any leakage so it wouldn't compromised the study. To ensure this a sealing test was done in all the cells. Guarantee the sterilization of the cell and the materials for the inoculation was also required so to ensure that every glass/plastic material went through an autoclave at 121°C and the cell (MFC) parts. Every single part from the MFC was sterilized using Ethanol 80% concentrated bath for 20-30 min and 30 min exposure to UV light. (Phillips, P: TUV 15WaH/ G15T8 ≈40cm)

For the MFC trials the following solutions were made using phosphate buffer instead of distilled water, this way the volume is slightly minimalized and therefore we can increase the volume of microorganisms, as represented in table 1.

S. marcescens was inoculated in 18 mL of LB solution with 2 mL of *S.m.*, for every trial the temperature for the inoculation was kept at 37°C in a controlled environment without agitation. The microorganism are added in the MFC after 23 hours of inoculation.

Table 1 - List of the MFC baseline tests and introduction of *S. marcescens*.

| Tests | Composition | Anode | Cathode |
|-------|---|-------|---------|
| 1 | Glucose (400 μ L) [1.5 M] | X | |
| | Potassium Ferricyanide (400 μ L) [0.02 M] | | X |
| 2 | Methyleme Blue (400 μ L) [0.01 M] | X | |
| | Potassium Ferricyanide (400 μ L) [0.02 M] | | X |
| 3 | Glucose (200 μ L) [1.5 M] + Methyleme Blue (200 μ L) [0.01 M] | X | |
| | Potassium Ferricyanide (400 μ L) [0.02 M] | | X |
| 4 | Glucose (100 μ L) [1.5 M] + Methyleme Blue (100 μ L) [0.01 M] + LB solution with <i>Serratia marcescens</i> (200 μ L) | X | |
| | Potassium Ferricyanide (400 μ L) [0.02 M] | | X |

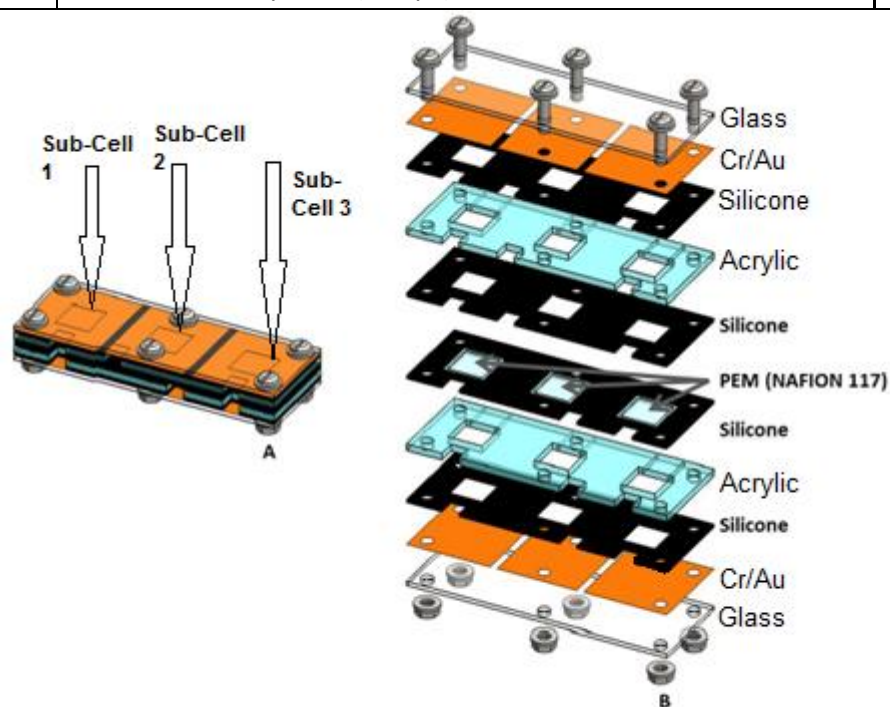


Figure 3 - Reference Microbial Fuel Cell structure adapted from (Fernandes, Rocha, & Carneiro, 2014).

Results and Discussion

1st Phase - 3D printed Microbial Fuel Cell

In this early state we'll start to see how the microorganism in study does behaves and how is this behavior being affected by the presence of new materials. Because the biocompatible properties in a microbial fuel cell and especially in the anode can define the adhesion of the microbes to the material. (Mustakeem, 2015)

In all the tests presented in this section, the growth of microorganisms was carried out in 75 ml of LB solution, inoculated with 7.5 ml of microorganisms. The growths were carried out with an agitation of 120 rpm and a constant temperature of 30 °C and 37°C in order to see the impact of the temperature in the growth process, and with the purpose of studying a controlled environment temperature and a temperature influenced by the season during the tests were being performed. The PLA trials were carried out at 37 °C and 120 rpm of agitation.

The cell mass of all the trials was measured by reading the absorbance at 600 nm as it can be seen in figure 4.

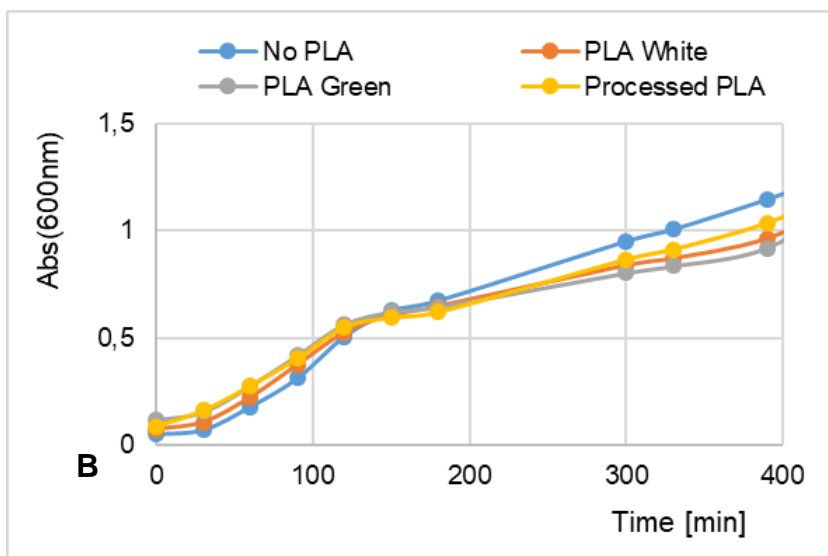
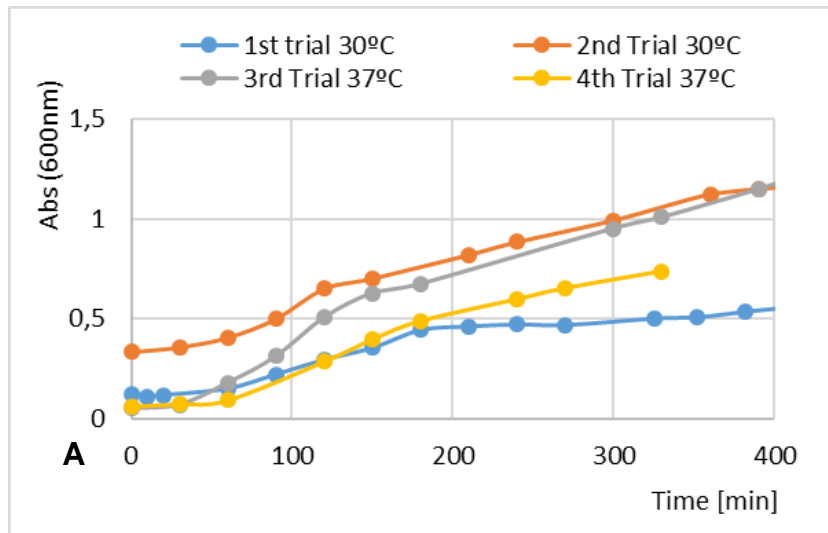
In figure 4A it's possible to see the *Serratia marcescens* growth curve for two difference temperatures (30 °C; 37 °C) this variance of temperature appears to have the same growth rate in general for the culture in study.

We can see in the figure 4B and 4C that the growth curve of our bacteria has the same behavior with and without de presence of the PLA. This means that we got good results concerning the new polymeric material, PLA, hence shows no negative impact to the *Serratia marcescens* growth rate.

A 3D sample was printed with these materials as compact as possible with very thin filaments to avoid any porosity. Though this 3D printed cell fails containing the liquids (H₂O) being only sustainable for 2 hours maximum after that time leakage would be perceived in the top cover parts and 5 hours later in the central (main container) part of the cell.

This only discards the 3D printed process used for this trial, the process itself delivers parts with a substantial porosity that doesn't allow the parts to be used on experiments involving liquids.

However, PLA is still validated as a possible material to be used on futures experiences with microorganisms due to not affecting its growth rate and for its resistance to microbial attack compared to other microbial and synthetic aliphatic polymers. The degradation of PLA is slow and that takes a long time for degradation to start. (Tokiwa, Calabia, Ugwu, & Aiba, 2009)



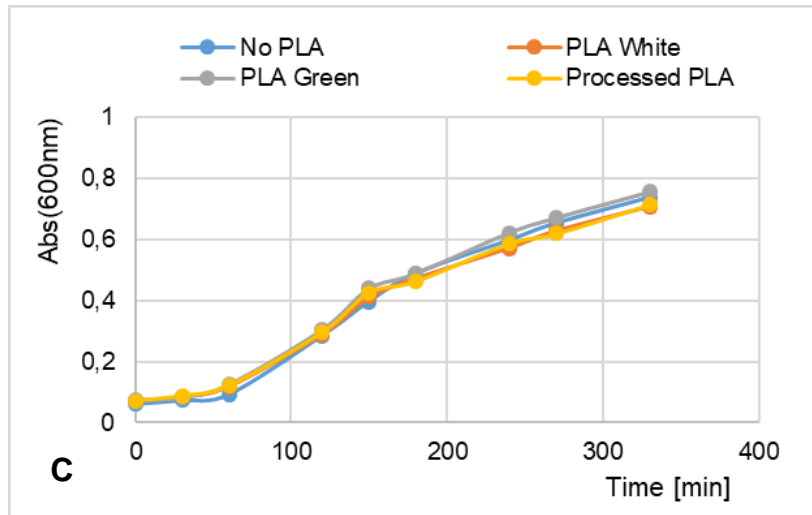


Figure 4 - Growth curve for *Serratia marcescens*. b) Presence of PLA trial 1; c) presence of PLA trial 2

2nd Phase – Conductor fabric as MFC electrode

In this part of the experiment the cell structure in use is similar to the one presented in figure 3 with a few changes, instead of the glass we have polycarbonate parts and in the conductive fabric goes to Cr/Au place. Before of each trial the structure and the conductive fabric went through the sterilization process mentioned in chapter 2 and in Appendix 1.

This structure contains the solutions perfectly which allowed for the research to proceed to the next step, conductivity trials using a [Cu+Ni+Au] conductive fabric as electrodes. The cells used from the MFC were the left one (1) and the one on the right (3) in all trials except for the ones in series.

The conductivity test was done with Phosphate Buffer and Glucose Figure 5a and 5b respectively. It's possible to see some electrical charge going throughout the time in case of glucose and in we can also see a peak of charge even when the same solution was being tested in both anodic and cathodic chambers. This another proof that our cell at this point is containing properly our solutions.

Moving on to what we call the test for baselines, to evaluate all the solutions isolated and together, compare them to the trial with *S.marcescens* and see their contribution and if the microbial component really optimizes our system. In figure 6 - 8 we can see the baseline trials 1 - 3 stated on Table 1 and in figure 9 a resume of the two best results from these baselines. Trial 4 presented on table 1 can be seen on figure 10.

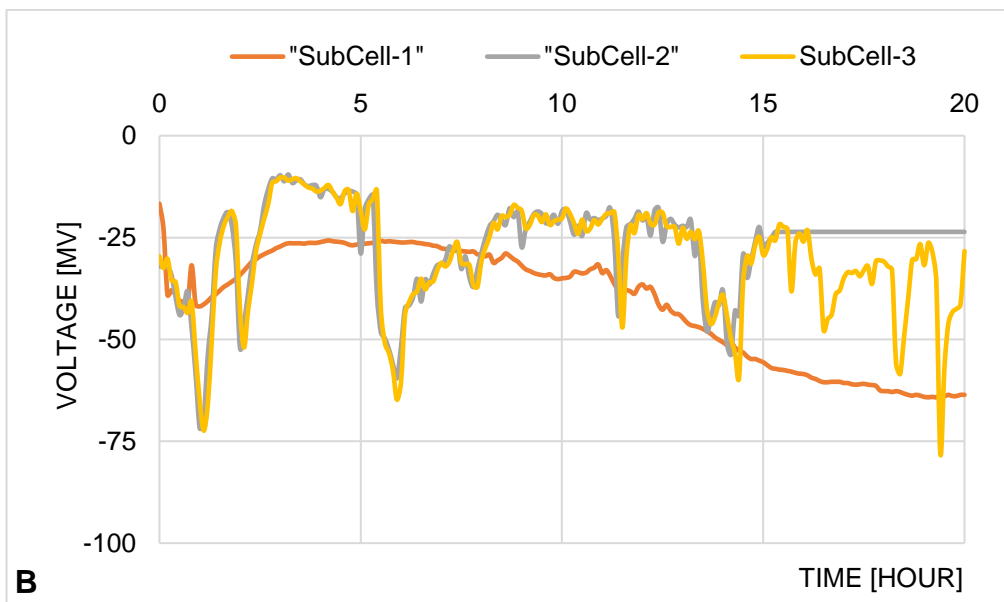
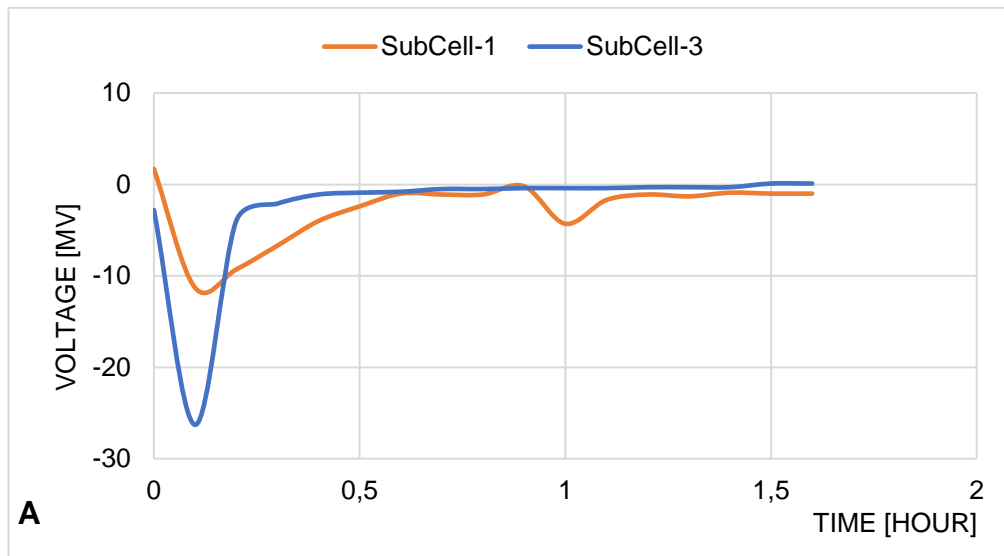


Figure 5 - a) Phosphate Buffer trial in two sub-cell compartments; b) Glucose trial in two sub-cell compartments

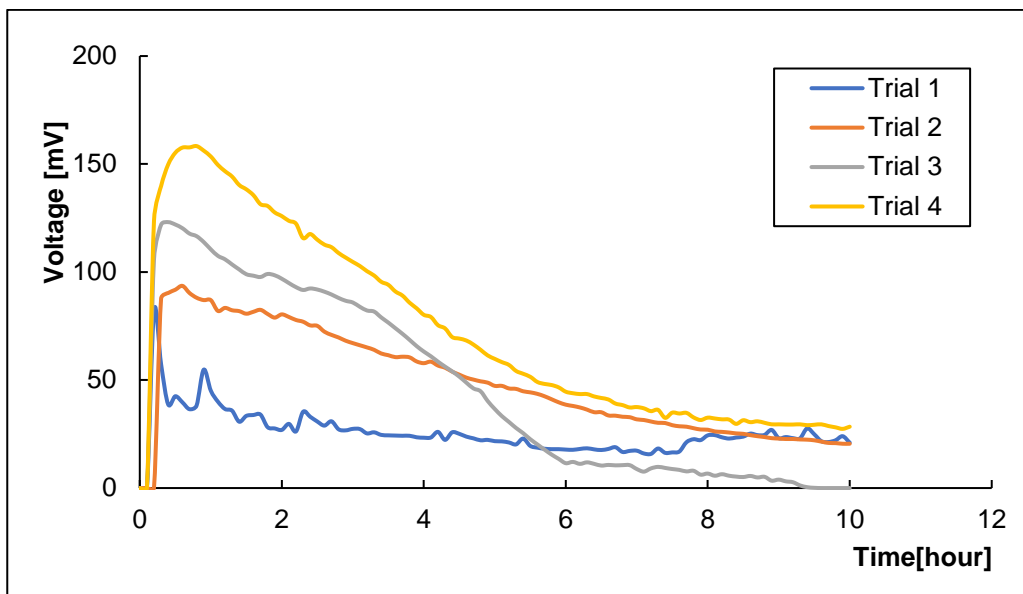


Figure 6 – Results for Glucose Baseline (1).

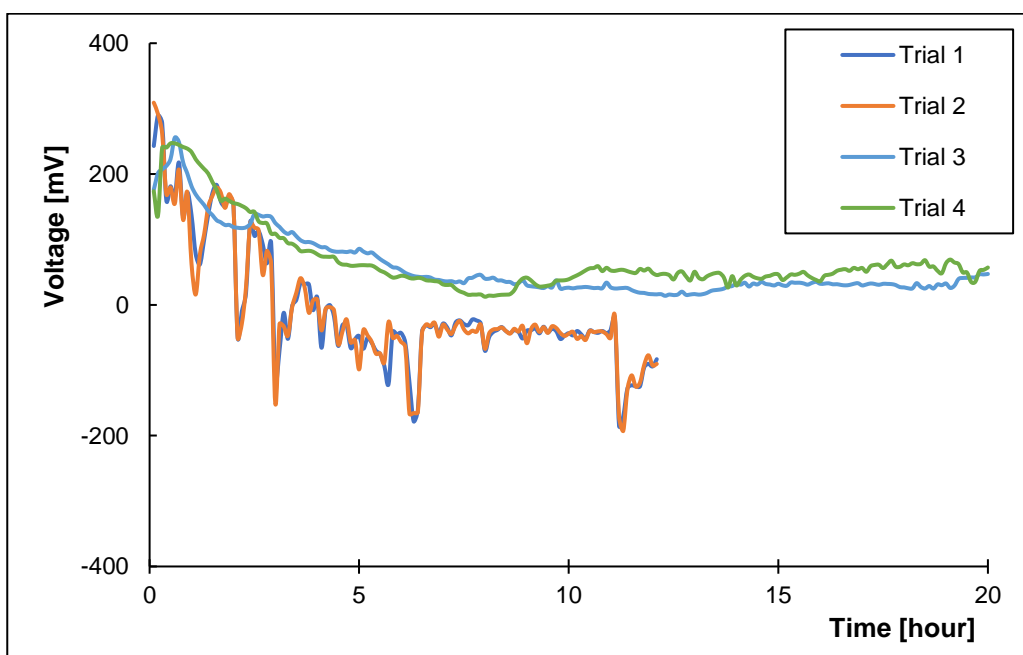


Figure 7 – Results for Methylene Blue Baseline (2).

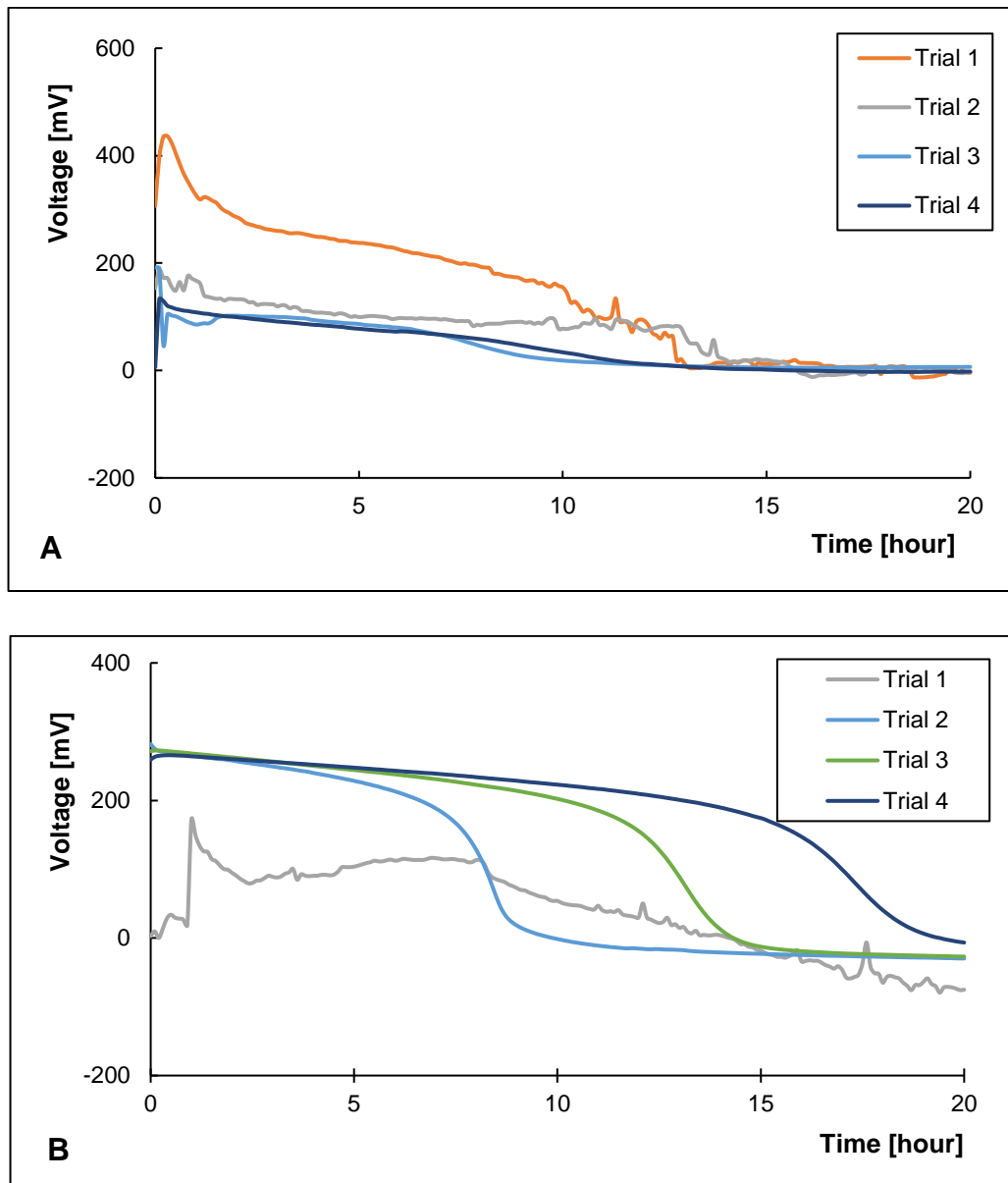


Figure 8 – Results for the combo Glucose + Methylene Blue Baseline (3); A – Dry-up conductive fabric; B – Conductive fabric previously bathed in Phosphate buffer solution.

When the conductive fabric is previously humidified has a certain stability for a longer period of time as we see in figure 8B.

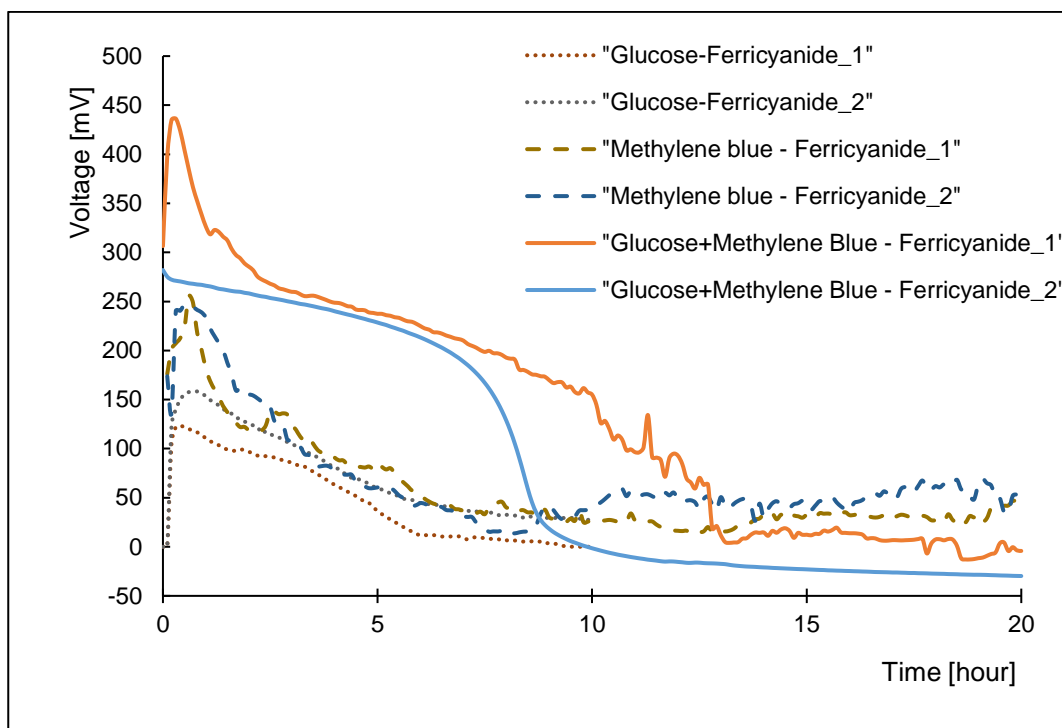


Figure 9 – All baselines, this graphic displays only the two best results from all the trials performed.

The results in figure 9 show a maximum of 436 mV that corresponds to 436 nA being produced by one single sub-cell of the MFC, since 1 M Ω resistor was being used. The peak is from the baseline 3, Glucose + Methylene Blue as the anode and Ferricyanide as the cathode.

Introducing *Serratia marcescens* to the trials

In all the tests presented in this section, the growth of microorganisms was carried out in 20 ml of LB solution, inoculated with 2 ml of microorganisms. The growths were carried out with a constant temperature of 37°C. The cell mass of all the trials was measured by reading the absorbance at 600 nm. In figure 10 we see a typical growth curve for our bacteria. We can detect two different behaviors, one we can see all the stages of the growth (lag, log and stationary) reaching the stationary stage 24 hours after the inoculation and the other one we see that the bacterial culture hasn't reach the stationary state even 30 hours after the inoculation. Compared to the study from (Fernandes, Rocha, & Carneiro, 2014) we already can spot a difference, somehow our inoculations were influenced between 5h – 25h that led to a different growth rate, most likely temperature fluctuations influenced by the environment where the culture was staying. In the previous study the best state for *Serratia marcescens* ended up to be the stationary state with an absorbance at 600 nm of ≈ 1.7 (14 hours after inoculation) and in this study the

stationary state is at an absorbance (600 nm) of ≈ 1.5 and in the present study is needed ≈ 23 hours for the bacteria to reach this state.

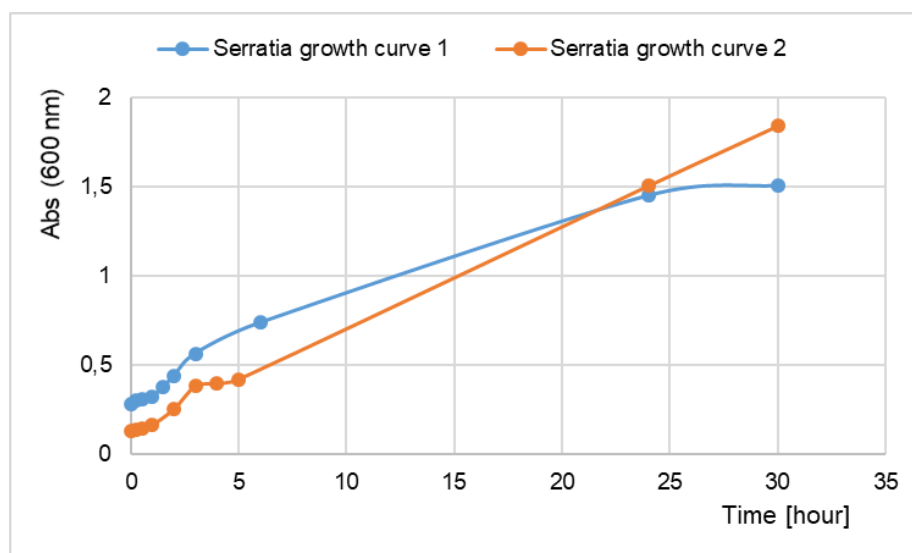


Figure 10 - *Serratia marcescens* growth curve with test conditions through 30 hours.

When adding the 100 μL of *S.marcescens* and reducing to half (100 μL) the Glucose volume, the electric potential registered doesn't go above the 200 mV, as seen in figure 10, we get one rehearsal with a continuous electric charge during approximately 14 hours and 2 other that have their peak in the first minutes and go immediately downwards in the next 4 hours.

The conducting fabric doesn't have the same electron acceptance as a nanocarbon based electrode or a graphene coated electrode would've, this and the possibility of the concentration of substrate in the anode chamber be insufficient for the number of microorganisms in the anode, can be one explanation for the results obtained. (Mustakeem, 2015) There are a lot of variables associated to this new microbial fuel cell technique, due to the alternative materials that has being used.

We can see in figure 10 that different concentration of *Serratia marcescens* show different behaviors on the MFC, the middle concentrated cultures of the four show the highest currents and for the first 4 hours we got an average current of 171.902 nA for the culture with 1.592 Abs (600 nm) and 117.8122 nA for the culture with 1.723 Abs (600 nm).

This difference shows that the inoculation process was suffering an external interference that made the process longer than it was theoretically expected to be and, therefore the trials in the MFC were not always performed in the stationary stage of the bacteria. This can explain part of the results that we see on figure 11. For the "Serratia 1.592 Abs" and "Serratia 1.723" we see periods of great energy production which indicates that

the inoculum used was indeed in the stationary stage, on the other hand the other two results indicates that most probably the bacteria was still on the logarithmic stage.

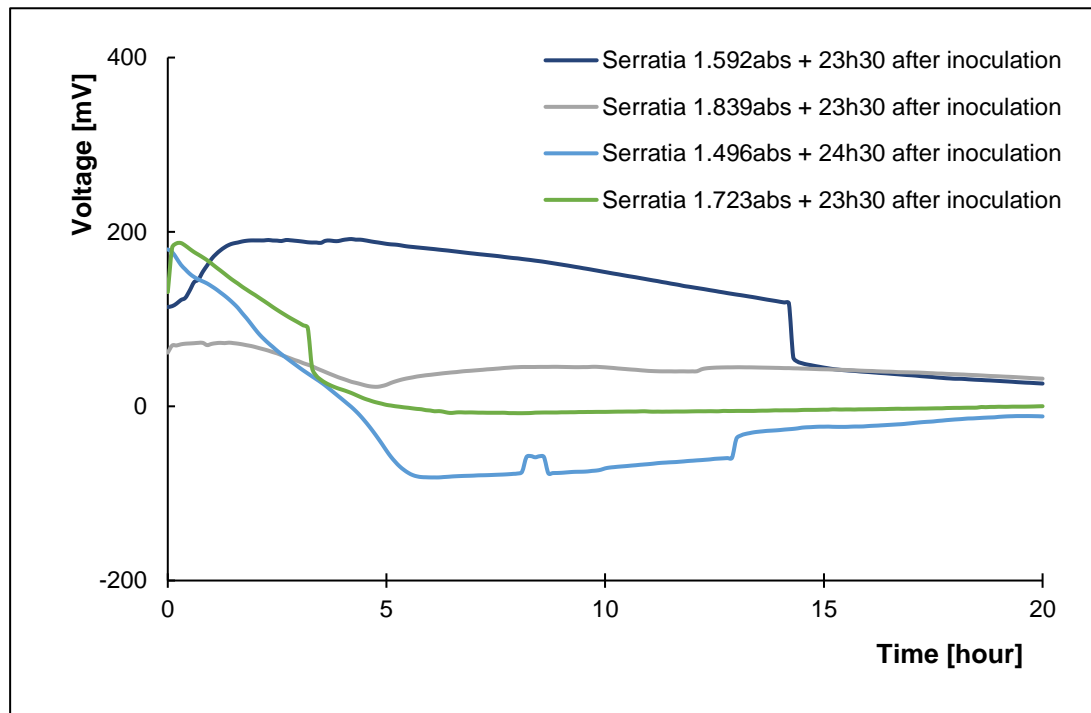


Figure 11 – Introduction of *Serratia marcescens* (4) in the MFC trials.

We will analyze the data for Glucose + Methylene Blue and after adding *S. marcescens* in the first 5 hours to see if we can find some similarity between them and see if the energy being produced is influenced by the *Serratia marcescens* or only the Glucose., for that we will make a brief statistic.

Statistical view

To better understand these results, it was performed an Analysis of Variance (ANOVA) between those 4 trials and see if we can find some statistical values that lead us to some conclusions about these tests.

Samples in study:

- A – *Serratia marcescens* 1.592 Abs (600 nm)
- B – *Serratia marcescens* 1.723 Abs (600 nm)
- C – Glucose + Methylene Blue – Potassium Ferricyanide _1
- D- Glucose + Methylene Blue – Potassium Ferricyanide _2

ANOVA is one of the most frequently used methods in statistics and it allows us to compare more than two group means in a continuous response variable, which is our case. To perform an ANOVA model we're assuming 3 things:

1. the probability distribution of responses in each group is normal.
2. Each probability distribution has the same variance.
3. Samples are independent.(Liu, 2015)

The null hypothesis for Welch's F-test is H_0 : "The means are equal"

The null hypothesis for Levene's test is H_0 : "The variance are equal"

Table 2 – Standard Statistics for the 4 populations in study.

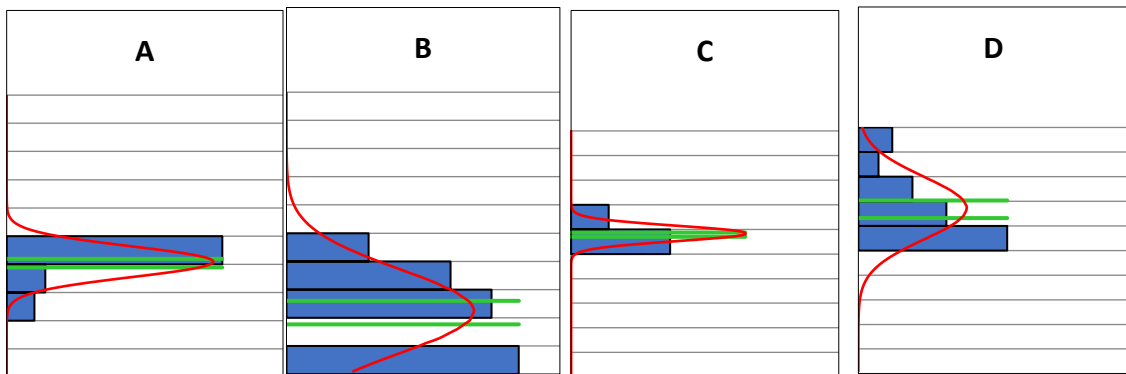
| SUMMARY STATISTICS | | | | |
|---------------------|----------|----------|----------|----------|
| | Sample A | Sample B | Sample C | Sample D |
| SAMPLE SIZE | 201 | 201 | 201 | 201 |
| MEAN | -36.313 | -56.820 | -7.251 | 128.704 |
| STDEV | 128.799 | 99.669 | 179.397 | 104.857 |
| SKEWNESS | 1.002 | 1.193 | 1.142 | -0.050 |
| KURTOSIS | -0.776 | 0.101 | -0.414 | -1.691 |
| MIN | -146.100 | -146.100 | -146.100 | -12.900 |
| 1ST QUARTILE | -130.800 | -130.800 | -130.800 | 12.400 |
| MEDIAN | -104.100 | -104.100 | -104.100 | 155.000 |
| 3RD QUARTILE | 113.800 | 1.500 | 237.400 | 233.300 |
| MAX | 191.800 | 187.200 | 436.300 | 282.000 |

Table 3 - Welch ANOVA results.

| WELCH ANOVA | | | | | |
|---|-----------------------|-------------------------------|---------------------|----------------|-----------------|
| HO : means are equal | Sum of Squares | Degrees of Freedom | Mean Squares | F ratio | P value |
| Among Groups | 1142637.447 | 3 | 380879,1491 | 224.9 | 7.85E-44 |
| Within Groups | 398253.1918 | 98.19 | 4055.944513 | | |
| Total | 1540890.639 | 203 | | | |
| H0 rejected, Group Means are different | | | | | |
| Overall statistics | | Test of equal Variance | | | |
| Grand Mean | 204.5926471 | Levene's test | | W critic | P value |
| Average Sample Size | 51 | statistic W | 40.13649422 | 2.649751643 | 2.39E-20 |
| Number of Groups | 4 | Variances are unequal | | | |

The red curve represents the estimated population distribution assuming normalilty

The green lines represent the 95% Individual Confidence Intervals for Means



Through ANOVA we can say with 95% confidence that there is an heterogeneity between the means and the variances of these 4 groups or at least one of them, since when the null hypothesis is rejected from different independent groups, ANOVA only provides information that the means of the four groups may differ and at least one group may show a difference (Kim, 2017). At the histograms presented in Table 2 we can have an indication that the data of our four samples possibly doesn't tend to a normal distribution therefore this test will not give us a conclusion free of errors concerning *S. marcescens*. Since the data is not normal instead of looking for the means or the variance it would be better to check the medians of the groups, for that we will perform a Kruskal-Wallis test. The Kruskal-Wallis test is a non-parametric test, which means that it does

not assume that the data came from a distribution that can be completely described by two parameters, mean and standard deviation (the way a normal distribution can), it is less sensitive to outliers, and the efficiency loss compared to ANOVA decreases for large samples. Since our results are not normal then this test can be done.

The null hypothesis for Kruskal-Wallis test is H_0 : "The medians are equal"

Table 4 - Kruskal-Wallis Result.

| Kruskal-Wallis test | |
|------------------------------------|----------|
| K | 211.54 |
| K critic χ^2 approximation | 7.81 |
| Pvalue χ^2 approximation | 1.35E-45 |
| H0 rejected, medians are different | |

The Kruskal-Wallis test states that the medians are different comparing all four populations, which means that we can say with 95% confidence that the trials performed with the presence of bacteria are not the same that the ones with baseline 3 which means that the MFC is functioning the same way with *Serratia marcescens*.

To better understand the negative impact that is seen in the MFC efficiency after *S.marcescens* some additional tests should've been made with brand new fabric to see the impact of the reutilization of it in this study, even though every material was sterilized the UV light can only reach the surface of each part, so the internal fibers of this electrode can only be cleaned with the ethanol bath and this can be one of the causes for our results. After concluding the additional tests it's important to retain the best method from that and make new trials with different glucose concentrations and possibly at different stages of the bacteria growth. This path will help us to reach the best conditions for these MFC using a textile electrode. Unfortunately, these tests were not conducted due to stock rupture in the supplier for this conductive fabric.

Microbial Fuel Cell with 3 Sub cells in series

The three sub cells of the Microbial Fuel Cell were filled with the respective anode and cathode composition and connected in series to a circuit with 1M Ω resistor.

The expected results would've been to have approximately three times more power coming out from the system, however the actual data is presented in figure 11, and the energy being collected from the system dropped from the initial tests with the bacteria.

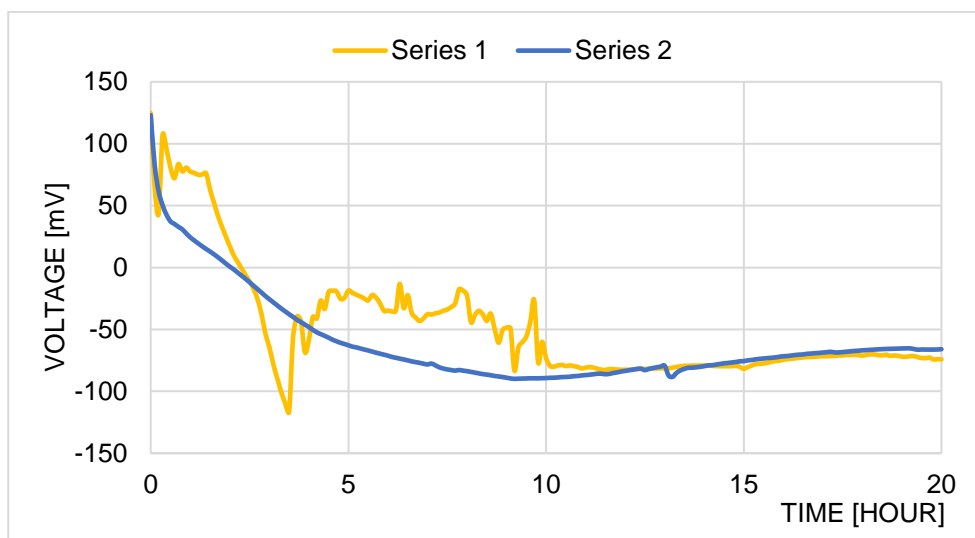


Figure 12 - MFC test in series with all three compartments connected to the circuit.

It's possible to see a peak of around 100 nA at the first half an hour and after that the current from the system drops to negative values a close indicative that there's no more substrate available in the anode or that our microorganisms are not feeding as it is supposed to.

In these later batches of *Serratia marcescens* the culture took at least 5 to 6 hour more to reach the absorption value at 600 nm than usual, the color was also a little lighter, this probably is one of the main reasons to a low energy production for the trial in series.

The conductive fabric had been reutilized since the introduction of bacterial culture, due to out of stock from the supplier and at these stages the electrode were too castoff due to all the process of sterilization, some of the fibers were already peeling out and the porosity was starting to be seen. The Microbial fuel cell was tightened even more too avoid any leakage of contaminating material though the use of the same cell also provoked the shatter of the Polycarbonate parts. I believe all that together led to the results presented in figure 9 and 10, because even though the fabric as an electrode isn't the most optimized choice, it proved to work and ended up to get 436nA as its maximum current without microbial influence and 191 nA with it.

The electrons released from microbes have to travel along an external circuit after passing through the anode, thus the conductivity of these materials is a determinant aspect for a study like this. The high electrical conductivity of the electrode material makes the electron flow with less resistance. At the same time, the interfacial impedance should be low to facilitate the electron transfer. At the cathode, the ionic conductivity is also required to facilitate triple phase boundary reaction. Therefore, the material for electrodes should be durable as well as stable in an acidic and a basic environment. (Mustakeem, 2015) And the conductive fabric in study presented to be durable for at

least 3 to 4 sterilization process and from then on each time it went through that process and through another microbial trial the fibers would wear out more and more.

Third phase – Graphene

The introduction of graphene as a new electrode, the procedure to be done would be similar, the structure used is presented in figure 13, and the concentrations for the anode and cathode components would be the ones stated in table 5, 6 and 7. Initially the purpose of this phase would be to combine the graphene with the [Cu+Ni+Au] conductive fabric and also use the reference (figure 3) construction adding graphene to the electrode Cr/Au, but the main supplier for [Cu+Ni+Au] conductive fabric stopped supplying to us.

First, we need to find new optimal conditions so, new concentrations were recalculated in order to improve the previous steps and to try and find the optimum condition for this specific design with these particular materials, and at the same time try to understand the balance and the exact contribution of the main solutions to the energetic capability of the microbial fuel cell.

In table 5 we keep the same conditions apart from the glucose volume to be used that is now 150 μL ; in table 6 we decrease the glucose concentration, keep the volume as 150 μL , double the concentration for Methylene Blue to 0.02 M and get half the concentration for Potassium Ferricyanide (0.01 M); in table 7 the glucose concentration goes back to 1.5 M and it's maintained the same conditions in table 6.

Table 5 - Proposed n°1 concentrations to reach optimal conditions.

| Solution | Molecular Formula | [] M | Volume (μL) in cell |
|--|---|-------|----------------------------------|
| Glucose | $\text{C}_6\text{H}_{12}\text{O}_6$ | 1.5 | 150 |
| Potassium Ferricyanide | $\text{C}_6\text{FeK}_3\text{N}_6$ | 0.02 | 400 |
| Methylene Blue | $\text{C}_{16}\text{H}_{18}\text{ClN}_3\text{S} \cdot x\text{H}_2\text{O}$ ($x=2-3$) | 0.01 | 50 |
| LB solution + <i>Serratia marcescens</i> Abs(600 nm)~1.7 | - | - | 200 |

Table 6 - Proposed n°2 concentrations to reach optimal conditions.

| Solution | Molecular Formula | [] M | Volume (uL) in cell |
|--|---|-------|---------------------|
| Glucose | C ₆ H ₁₂ O ₆ | 1 | 150 |
| Potassium Ferricyanide | C ₆ FeK ₃ N ₆ | 0.01 | 400 |
| Methylene Blue | C ₁₆ H ₁₈ CIN ₃ S xH ₂ O (x=2-3) | 0.02 | 50 |
| LB solution + <i>Serratia marcescens</i> Abs(600 nm)~1.7 | - | - | 200 |

Table 7 - Proposed n°3 concentrations to reach optimal conditions.

| Solution | Molecular Formula | [] M | Volume (uL) in cell |
|--|---|-------|---------------------|
| Glucose | C ₆ H ₁₂ O ₆ | 1.5 | 150 |
| Potassium Ferricyanide | C ₆ FeK ₃ N ₆ | 0.01 | 400 |
| Methylene Blue | C ₁₆ H ₁₈ CIN ₃ S xH ₂ O (x=2-3) | 0.02 | 50 |
| LB solution + <i>Serratia marcescens</i> Abs(600 nm)~1.7 | - | - | 200 |

With this new set of experiments we have a new baseline for this new MFC using graphene and we can from these results set the best concentrations and volumes. It would also be interesting to test different concentrations *S. marcescens* between 1.4 Abs (600 nm) and 1.9 Abs (600 nm) just to see if there is actually a better range at this microorganism state to increase their proliferation inside the MFC and therefore contribute to more free electrons being released.

In this stage, to avoid more variables the structure that is going to be used just like the referenced cell (figure 3) but instead of the carbon nanotubes we'll have a graphene film deposited on the same electrical conductive material (Chromium and Gold). For the Cr/Au deposition the procedure used was the Physical vapor deposition also known as PVD on a thinner glass (this glass was previously machined with a diamond tip drill in order to perform the screws holes without breaking the entire glass).

Table 8 - Reference characteristics used for the Cr/Au film deposition in this study. (Fernandes, Rocha, & Carneiro, 2014)

| <i>Characteristics</i> | <i>Chromium</i> | <i>Gold</i> |
|-------------------------|---------------------------|---------------------------|
| <i>Z-factor</i> | 0.305 | 0.381 |
| <i>D</i> | 7.2 gm/ cm ³ | 19.2 gm/ cm ³ |
| <i>Initial Pressure</i> | 3.2x10 ⁻⁶ mbar | 2.4x10 ⁻⁶ mbar |
| <i>Final Pressure</i> | | 4.0x10 ⁻⁶ mbar |
| <i>Current</i> | 20-30 mA | 120 A |
| <i>Voltage</i> | 10 kV | 1.3 V |
| <i>Deposition rate</i> | ≈ 120 Å/s | ≈ 3.1 Å/s |
| <i>Thickness</i> | 120 nm | 200 nm |
| <i>Technique</i> | PVD E-beam | PVD Resistive Evaporation |

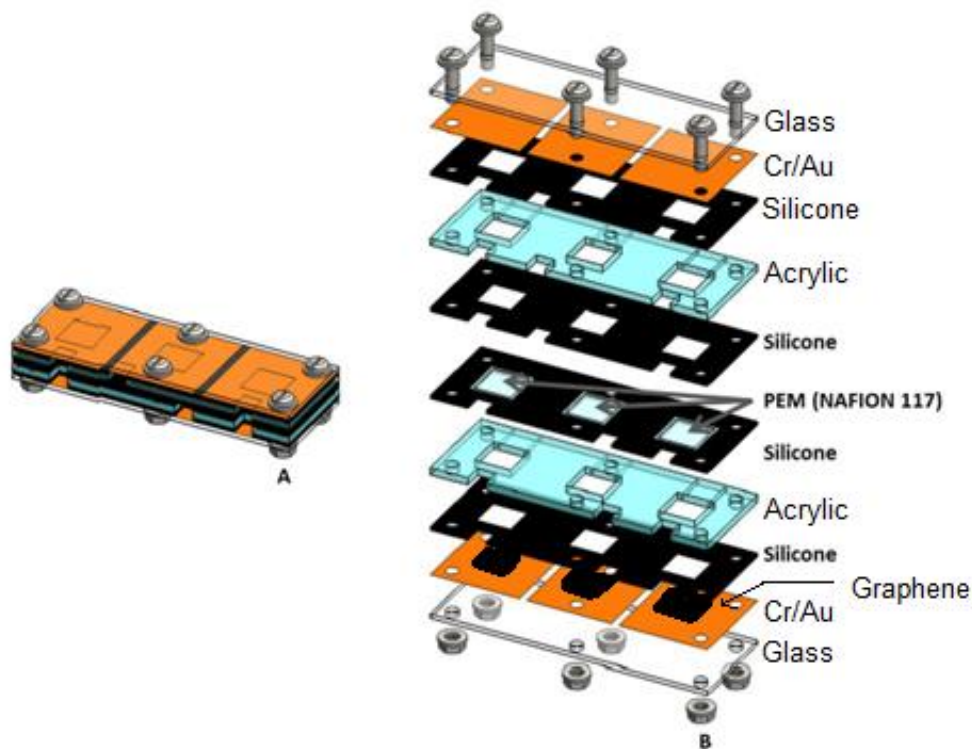


Figure 13 - MFC structure with graphene as an electrode. Adapted (Fernandes, Rocha, & Carneiro, 2014).

Graphene has been attracting strong scientific and technological interest with great application potentials in various fields, such as lithium ion batteries, solar cells and electrochemical super-capacitors, for its unique nanostructure and extraordinary properties

such as high surface area that allows the capture of electrons that come from all direction in comparison with NTC, excellent conductivity, outstanding mechanical strength and extraordinary electro catalytic activities. (Zhang., 2011)

Using graphene in the MFC and the best conditions in the anodic chamber we not only are optimizing the electrical source but also the collection of the energy that is being produced on the cell, because the graphene film on top of the Cr/Au will increase the electron acceptance area and condition, hence a higher energy gain for our microbial fuel cell.

Unfortunately, of graphene was not possible to proceed due to a not yet optimized procedure for the graphene deposition in our facilities. But the studies stay open for when the chance to proceed with it appears.

4

Conclusion

The outcomes of this thesis were not what we initially expected for this research, though we were trying to test a new set of materials and in science we cannot always expect to have positive results, but we are always learning something new and look forward to try new methods, techniques and materials.

Despite the unavailability of some of the materials that didn't allow the research to continue, that are some positive conclusions to attain from this study:

- The presence of Polylactic Acid (PLA) doesn't not affect the normal growth rate of *Serratia marcescens* and that this strain is actually pretty resistant;
- Biodegradable materials such as PLA are a possible material to be use in future studies involving pathological strain of bacteria;
- The [Cu+Ni+Au] conductive fabric can be used as a capable electrode for MFC studies, though the architecture implemented in this study isn't the best for this type of electrode;
- This research show that [Cu+Ni+Au] conductive fabric does have its limitations should not be used too many consecutive times but, it still allowed to recollect some of the energy being produced in the MFC.
- Compared to our referenced study the bacteria didn't always had a stable growth state and the stationary stage oscillated depending on external factors, so a better knowledge of *Serratia m.* growth should've been done in order to find the external factor and exactly at what time (after inoculation) should the bacteria reached its stationary state. In the reference study we had a mark of aprox. 14h to reach the optimum stage and in this study we needed aprox. 23h in some cases and more than 30h on other cases.
- In this study the peak of current that was able to get with a full MFC working structures was aprox. 200 nA for about 5 consecutive hours, was lower than previous reference study where was possible to generate 616 nA. Though is important to enhance that in this study was not used carbon nanotubes vertically aligned as it was used on the reference study and that contributed to a better capture of the electrons within the cell.

In the future, probably the reutilization of the material would be something to lose and try and use always brand-new fabric to not attack the fibers with the aggressive process of sterilization nor the possible contamination of the bacterial culture. Probably a new cell design would be beneficial in future studies using conductive fabric, this one had the contact area to the circuit on the bottom and to avoid possible leaks through the fibers a MFC with the electrodes coming from above would be the best choice.

Future Perspectives

I believe that Microbial Fuel Cells studies will be an advantage to science and the world. With too much fossil fuel consumption and the limitations of renewable energy, is imperative that an alternative may be found.

Combining the waste waters around the globe, or starting to power up small electronics with these type of alternative energy using glucose or sucrose substrate, will have a gigantic impact in a good way, decreasing the human footprint and will help long term to save some of the precious land resources.

A fusion with a new and sprouting technology that we are creating on the daily basis, 3D printing, conductive materials made of textiles, photolithography, paper engineering are some of the examples.

In (Fraivan, Mukherjee, Sundermier, Lee, Choi, 2013) work, a paper-based MFC was created featuring a paper-based PEM and chambers for the anode and cathode. Flexible carbon clothes were employed as the electrodes and hydrophobic barriers in paper and carbon clothes were patterned with photoresist. This MFC was able to generate a current of 74 μA and an operating voltage of 74mV. This paper-based MFC has the advantages of being easy to use, are disposable and cost-effective, and a good solution for a power source for disposable diagnostic devices.

A new mini-MFC array was designed and microfabricated, and its electricity generation performance was evaluated. Compared to the previous mini-MFCs reported in the literature, the mini-MFC fabricated in this work has the highest maximum power density of the reported micro-liter scale MFCs and maximum current density of 29mW/m² and 2148mA/m², respectively. The microfabrication process, configuration design and utilization of air cathode are likely to have contribution to the great performance of the mini-MFC.

Another example is towards fabrication of small-sized MFCs for their potential application as miniature devices. A miniature volume and a low substrate consumption, makes a mini-MFC especially useful in medical, communication and other areas, and is also suitable for evaluating the electricity generation characteristics of microorganisms, such as electrochemical activity, maximal potential, and power density.

In (Chen, Zhao, Qiu, Chu, ..., (2010) work, a new single-chamber mini-MFC was fabricated using a micro fabrication technique by applying sputtering deposition, photolithography and ion-beam etching. A mini-MFC array was designed and micro fabricated,

and its electricity generation performance was evaluated. The mini-MFC fabricated in this work reported a very high power and current density of 29mW/m² and 2148mA/m², respectively.

There are also some frontline researches in the mutagenesis and even recombinant DNA technology that can possibly be used in the future to obtain some super microorganisms for MFCs. Microbes may be used as a pure culture or a mixed culture forming a synergistic microbial consortium to offer better performance.

One type of bacterium in a consortium may provide electron mediators that are used by another type of bacterium to transport electrons more efficiently to an anode (Rabaey & Verstraete, 2005), but it is possible that an optimized microbial consortium can be obtained to operate an MFC without extraneous mediators or biofilms while achieving superior mass transfer and electron transfer rates and this is also an innovating technique to be explored in the future (Du, Li, & Gu, 2007).

References

- Du, Z., Li, H., & Gu, T. (2007). A state of the art review on microbial fuel cells: A promising technology for wastewater treatment and bioenergy. *Biotechnology Advances*, 25(5), 464–482. <https://doi.org/10.1016/j.biotechadv.2007.05.004>
- Fernandes, M., Rocha, L., & Carneiro, C. (2014). Development of a Micro-Microbial Fuel Cell with Nanoimproved Electrodes.
- Ghanapriya, K., Rana, S., & Kalaichelvan, P. T. (2010). Electricity Generation from *Serratia marcescens* Isolated from Aerobic Sludge using Microbial Fuel Cell Technology and its Optimization. *Electricity Generation from Serratia Marcescens Isolated from Aerobic Sludge Using Microbial Fuel Cell Technology and Its Optimization*, 09(10), 32–38. Retrieved from http://www.advancedbiotech.in/archives_apr_10_Electricity.html
- Hejazi, A., & Falkiner, F. R. (1997). *Serratia marcescens*. *J. Med. Microbiol*, 46(1997), 903–912. <https://doi.org/10.1099/00222615-46-11-903>
- Kalathil, S., Patil, S. A., & Pant, D. (2017). *Microbial Fuel Cells: Electrode Materials*. Reference Module in Chemistry, Molecular Sciences and Chemical Engineering. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-409547-2.13459-6>
- Kim, T. K. (2017). Understanding one-way anova using conceptual figures. *Korean Journal of Anesthesiology*, 70(1), 22–26. <https://doi.org/10.4097/kjae.2017.70.1.22>
- Liu, H. (2015). Comparing Welch ' s ANOVA , a Kruskal-Wallis test and traditional ANOVA in case of Heterogeneity of Variance.
- Logan, B. E., Hamelers, B., Rozendal, R., Schröder, U., Keller, J., Freguia, S., ... Rabaey, K. (2006). Microbial fuel cells: Methodology and technology. *Environmental Science and Technology*, 40(17), 5181–5192. <https://doi.org/10.1021/es0605016>
- Lovley, D. R. (2006). Microbial fuel cells: novel microbial physiologies and engineering approaches. *Current Opinion in Biotechnology*, 17(3), 327–332. <https://doi.org/10.1016/j.copbio.2006.04.006>
- Mustakeem. (2015). Electrode materials for microbial fuel cells: Nanomaterial approach. *Materials for Renewable and Sustainable Energy*, 4(4), 1–11. <https://doi.org/10.1007/s40243-015-0063-8>
- Pant, D., Van Bogaert, G., Diels, L., & Vanbroekhoven, K. (2010). A review of the substrates used in microbial fuel cells (MFCs) for sustainable energy production. *Bioresource Technology*, 101(6), 1533–1543. <https://doi.org/10.1016/j.biortech.2009.10.017>
- Qian, F., Baum, M., Gu, Q., & Morse, D. E. (2009). A 1.5 microL microbial fuel cell for on-chip bioelectricity generation. *Lab on a Chip*, 9(21), 3076–3081. <https://doi.org/10.1039/b910586g>
- Tokiwa, Y., Calabia, B. P., Ugwu, C. U., & Aiba, S. (2009). Biodegradability of Plastics Bio-plastics, 3722–3742. <https://doi.org/10.3390/ijms10093722>
- Zhang, Y., Mo, G., Li, X., Zhang, W., Zhang, J., Ye, J., ... Yu, C. (2011). A graphene modified anode to improve the performance of microbial fuel cells. *Journal of Power Sources*, 196(13), 5402–5407. <https://doi.org/10.1016/j.jpowsour.2011.02.067>
- LIU, W., CHENG, S. (2014). Review: Microbial fuel cells for energy production from wastewaters: the way toward practical application. *Journal of Zhejiang University-SCIENCE A (Applied Physics & Engineering)*, 841–861.
- Kim, B., Chang, I., Gil, G., Park, H., & Kim, H. (2003). Novel BOD (biological oxygen demand) sensor using mediator-less microbial fuel cell. *Biotechnology letters*, 25(7):541–545, ISSN 0141-5492. PMID: 12882142.

- Chang, I., Jang, J., Gil, G., Kim, M., Kim, H. (2004) Continuous determination of biochemical oxygen demand using microbial fuel cell type biosensor. *Biosensors and Bioelectronics*, 19(6):607–613. ISSN 09565663. <http://linkinghub.elsevier.com/retrieve/pii/S0956566303002720>.
- Rittmann, B.E., (2008). Opportunities for renewable bioenergy using microorganisms. *Biotechnology and Bioengineering*, 100(2):203-212. [doi:10.1002/bit.21875]
- Liu, H., Grot, S., Logan, B.E. (2005). Electrochemically Assisted Microbial Production of Hydrogen from Acetate. *Environmental Science & Technology*, 39(11):4317–4320, ISSN 0013-936X, 1520-5851. <http://pubs.acs.org/doi/abs/10.1021/es050244p>.
- Fraiwani, A., Mukherjee, S., Sundermier, S., Lee, H.S., Choi, S. (2013) A paper-based microbial fuel cell: Instant battery for disposable diagnostic devices. *Biosensors and Bioelectronics*, 49: 410 - 414. <https://doi.org/10.1016/j.bios.2013.06.001>.
- Chen, Y.P., Zhao, Y., Qiu, K.Q., Chu, J., Lu, R., Sun, M., Liu, X.W., Sheng, G.P., Yu, H.Q., Chen, J., Li, W.J., Liu, G., Tian, Y.C., Xiong, Y. (2010). An innovative miniature microbial fuel cell fabricated using photolithography. *Biosensors and Bioelectronics*, 26: 2841-2846. <https://doi.org/10.1016/j.bios.2010.11.016>.
- Rabaey, K., Clauwaert, P., Aelterman, P., Verstraete, W. Tubular microbial fuel cells for efficient electricity generation. *Environmental Science and Technology* 2005, 39: 8077–82.

Appendix 1

Preparation of LB solution

For V= 250 mL of LB solution:

Reagents:

- Peptone – 2. 5 g
- Yeast Extract – 1. 25 g
- NaCl – 1. 25 g
- Distilled H₂O – till 250 ml

Preparation: Balanced every single reagent mixed all together and start dissolving with a little distilled water when all the solution is dissolved and presents no solids, fill the rest of the Shott flask till 250 mL total. Seal it and proceed to sterilization of the mean (auto-clave).

Biocompatibility - test method

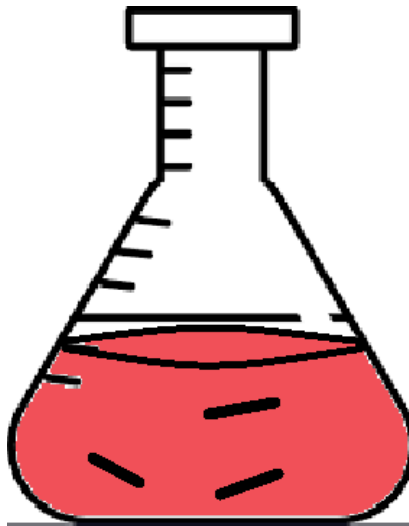


Figure 14 - Sketch of the PLA/S.m. biocompatibility test (Erlen Mayer with *S. marcescens*).

The unprocessed and processed PLA parts (previously sterilized) would be put inside the *Serratia marcescens* culture each material had its own cultures and it would be introduced 3 – 4 pieces of about 2-3 cm for the unprocessed material and 5 – 6 parts of 1 cm x 1 cm squares for the processed material

Sterilization process:

20-30 min – Bath of 80% concentrated Ethanol solution.

30 min – Exposed to UV light to surface sterilization.

Involved the parts with the silver paper also UV sterilized and keep the parts there till it's time do use.

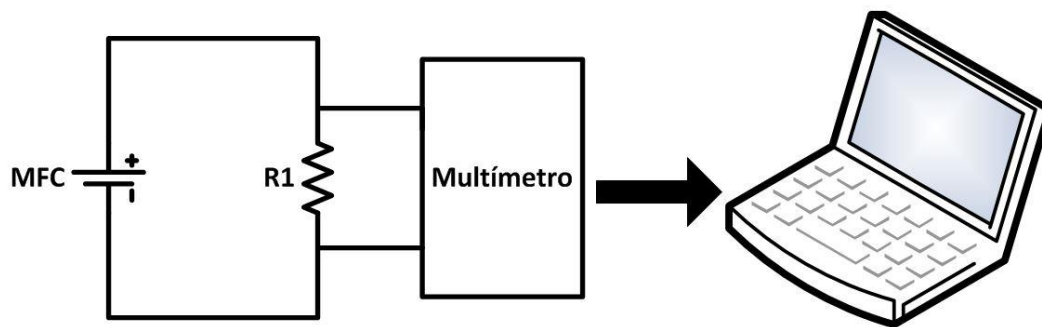
Microbial Fuel Cell Setup:

Figure 15 - Standard setup used for all the MFC tests using a resistor of 1 M Ω .